Immunomodulatory potential of gut microbiome-derived short-chain fatty acids (SCFAs)

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Intestinal microbiota is an element of the bacterial ecosystem in all mammalian organisms. These microorganisms play a very important part in the development, functioning, and modulation of the immune system from the moment of birth. In recent years, owing to the use of modern sequencing techniques, the microbiome composition in healthy people has been identified based on bacterial 16S rRNA analysis. Currently, more and more attention is being given to the influence of microorganisms on the host’s cellular metabolism. Analysis of microbial metabolites, among them short-chain fatty acids (SCFAs), and disruption of intestinal microbiota homeostasis in terms of their effects on molecular regulatory mechanisms of immune reactions will surely improve the understanding of the etiology of many common diseases. SCFAs, mainly butyrate, propionate, and acetate, occur in specific amounts, and their proportions can change, depending on the diet, age, and diseases. The levels of SCFAs are substantially influenced by the ratio of commensal intestinal bacteria, the disturbance of which (dysbiosis) can lead to a disproportion between the SCFAs produced. SCFAs are regarded as mediators in the communication between the intestinal microbiome and the immune system. The signal they produce is transferred, among others, in immune cells via free fatty acid receptors (FFARs), which belong to the family of G protein-coupled receptors (GPCRs). It has been also confirmed that SCFAs inhibit the activity of histone deacetylase (HDAC) – an enzyme involved in post-translational modifications, namely the process of deacetylation and, what is new, the process of histone crotonylation. These properties of SCFAs have an effect on their immunomodulatory potential i.e. maintaining the anti/pro-inflammatory balance. SCFAs act not only locally in the intestines colonized by commensal bacteria, but also influence the intestinal immune cells, and modulate immune response by multi-protein inflammasome complexes. SCFAs have been confirmed to contribute to the maintenance of the immune homeostasis of the urinary system (kidneys), respiratory system (lungs), central nervous system, and the sight organ.

Key words: intestinal microbiota, dysbiosis, short-chain fatty acids (SCFAs), inflammation, inflammasome

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INTRODUCTION

As reviewed by Mackowiak (Mackowiak, 2013), studies of the role that intestinal microflora plays in maintaining the organism’s homeostasis have been carried out for more than one hundred years. The century-long research on the human microbiome has confirmed that commensal bacteria found in various organs and integuments of the human body promote the pathogenesis of many diseases (Aragón et al., 2018; Evsutina et al., 2017; Flemer et al., 2017; Porter et al., 2018).

The development and composition of the human gut microbiota are influenced by the type of labor and infant nutrition (Nagpal et al., 2016). In healthy neonates, the intestinal microflora changes from birth to the age of three. Initially, it mainly consists of facultative anaerobic bacteria from the Enterobacteriaceae family, Enterococcus, and Staphylococcus genera. The composition of this ecosystem alters as a result of children’s growing up, the introduction of new foods in their diet, and exposure to environmental factors. Thus, when they reach the age of three, their intestinal microflora is represented by obligatory anaerobes, mostly Clostridium (C.) leptum, C. cocoides, Bifidobacterium, and Bacteroides fragilis (Nagpal et al., 2017).

At present, methods for deep sequencing of bacterial genetic material from feces samples had let us establish which bacteria commonly occur in the intestines, irrespective of individual features, past diseases, diet, and environmental factors. The available studies show that each person has their stable bacterial metagenome—the collective genome of microorganisms from the gut environment that provides information on the microbiological diversity and ecology of this specific environment (niche). As reviewed by Maynarda and others (Maynard et al., 2012) and Iebba and others (Iebba et al., 2016), the presence of diverse but balanced intestinal microflora is defined as eubiosis, which is characterized by a predominance of beneficial microbiota species, mainly belonging to two phyla—Bacteroidetes and Firmicutes. Although pathogenic species may be present, they are in the minority. Moreover, in the eubiotic state, the immune homeostasis of the intestinal mucous membrane and the gut-associated lymphoid tissue (GALT) is maintained. This is so because microorganism-associated molecular patterns (MAMPs) stimulate secretion of cytokines in-
volved in the differentiation of dendritic cells and macrophages, which results in the activation of Treg cells. Their activity manifests as secretion of IL-10, involved in the development of an anti-inflammatory environment. Additionally, transforming growth factor-β (TGF-β), also secreted by Treg lymphocytes, stimulates plasma cells to secrete immunoglobulin A (IgA). Individual metagenomes (mainly including bacteria from the Bacteroidetes and Firmicutes phyla) may slightly deviate from the one shared by the whole population. The presence of such a specific and stable microbiome ensures the smooth functioning of the whole intestinal ecosystem, undergoing modifications throughout the adult life (Qin et al., 2010). The intestinal balance between Bacteroidetes and Firmicutes may not be a defining attribute of an individual's intestinal microbiome. Instead, individuals have their own 'personal equilibrium' between Bacteroidetes and temporarily variable Firmicutes (Lloyd-Price et al., 2017; Claesson et al., 2011). Composition of the human intestinal microbiota has an effect on the maintenance of health, but its role in many pathological states is also taken into consideration (Holmes et al., 2012). Microflora influences the host's health through microbial components, such as polysaccharide A (PSA) (Mazmanian et al., 2008), for myl peptides (Bufe et al., 2015), D-glycero-β-D-manno heptose-1,7-bisphosphate (HBP) bacterial metabolites (Gaudet et al., 2015), and metabolites derived from nutrients with the help of bacteria, namely vitamins, amino acids, metabolites that are ligand for aryl hydrocarbon receptor (AHR) (Jin et al., 2014; Natividad et al., 2018), polyamines (Hesterberg, Cleveland, & Epling-Burnette, 2018), and short-chain fatty acids (Gill et al., 2018).

**INTESTINAL DYSBIOSIS**

Microbiome profiling in healthy people provides data on the microbiota in particular organs. Individual differences in the human microbiome are specific, functionally important, and personalized (Human Microbiome Project Consortium, 2012). It has been also noticed that Bacteroides prevail in people with small variety of Firmicutes (Human Microbiome Project Consortium, 2012).

Balance between organisms making up the intestinal ecosystem may be disrupted not only by taking broad-spectrum antibiotics, but also by antiviral or antifungal drugs. Changes in human gut microbiota composition can be associated with commonly used non-antibiotic drugs, such as proton pump inhibitors (PPIs), nonsteroidal anti-inflammatory drugs (NSAIDs), atypical antipsychotics (AAPs), but also by antibiotics – metformin (Maier et al., 2018). The modulation of the intestinal microbiota composition is influenced by the variety in diet, long-lasting stress, and adverse environmental factors (among them bisphenol A) (Malaíse et al., 2017).

Imbalance between microorganisms naturally colonizing the host, and their disturbed functioning, is called dysbiosis. In macroorganisms inhabited by microorganisms, such a situation triggers the immune response, affects the host’s metabolism, and can possibly contribute to the development of inflammatory diseases (Lee & Kim, 2017; Miyoshi et al., 2017; Saltzman et al., 2018; Slingerland, Schwabkey et al., 2017; Zechner, 2017).

Owing to the fact that different groups of microorganisms are involved and interact with each other, several types of dysbiosis can be distinguished. One example is the presence of pathobionts – symbiotic microorganisms that under normal circumstances do not affect the host’s health and do not elicit an inflammatory response. But under particular conditions (environmentally induced) they have the potential to cause dysregulated inflammation leading to a disease (Chow et al., 2011). Moreover, pathobionts are capable to expanding during episodes of inflammation and may exacerbate the disease process. For example Clostridium (C.) difficile, C. perfringens, Fusobacterium nucleatum or enterotoxigenic strain Bacteroides fragilis, occur in the healthy human gut microbiome (Maier et al., 2018), but in some cases are considered as pathological factors (Fukugita et al., 2015; Komiyama et al., 2018; Purcell et al., 2017; Tajkarimi & Wexler, 2017). Moreover, in the murine IBD (inflammatory bowel disease) model, it was shown that a pathobiont (Helicobacter bilis) may synergize with commensal strains of bacterial microbiota and exacerbate pathology in this disease, which was demonstrated by the increase in TGFβ and proinflammatory cytokines (IL-1β, IL-17A, IL-17F), secreted by the Th17 cells (Gomes-Neto et al., 2017). Nevertheless, as a result of gene mutation in bacterial type VI secretion system (T6SSs) or intestinal factors, such bacteria may cause infections and inflammatory states in mice (Chow & Mazmanian, 2010; Maier et al., 2018). Another example of a pathobiont is Helicobacter (H.) hepaticus, which can be involved in inflammatory gut diseases through the induction of Th17 proinflammatory lymphocytes. A murine model-based research (Xu et al., 2018) demonstrated that inactivation of c-MAF transcription factor in Treg lymphocytes disturbed their differentiation and function, which was manifested by lower IL-10 levels. This, in turn, favored accumulation of the Th17 cells and increased inflammatory response to H. hepaticus (Xu et al., 2018).

Another contributor to dysbiosis is the loss of commensal bacteria, stemming from elimination of bacterial cells, or inhibition of their proliferation (Korem et al., 2015).

Dysbiosis is also characterized by lower diversity of bacterial species inhabiting particular places in the body. Diversity of commensal microorganisms is shaped from birth (Nagpal et al., 2017). What is more, it can be modulated by dietary habits (Cotillard et al., 2013), as well as obesity and related metabolic changes (Le Chatelier et al., 2013).

On the basis of a murine model, it has been demonstrated recently that the intestinal microbiota are also affected by the process of aging (Langhans et al., 2008), which can contribute to systemic inflammatory states (inflammaging) (Kim et al., 2016; Fransen et al., 2017). The reported results suggest that ‘old’ intestinal microbiota transplanted to germ free (GF) mice increase the production of tumor necrosis factor α (TNF-α), whose chronic secretion can exert adverse effects on metabolic processes and insulin resistance, and produce inflammatory changes in the arterial walls. Furthermore, ‘aging’ of the intestinal microbiota was proved to be a far more important source of proinflammatory bacterial factors (including bacterial lipopolysaccharide, LPS) released to the blood circulation system, than the microbiota of young animals. It was also found that with age, Firmicutes outnumber Bacteroidetes (Kim et al., 2016; Fransen et al., 2017). A human population based study (Claesson et al., 2011) demonstrated that the intestinal microbiota phylum predominating in aging individuals is Bacteroidetes. In younger adults, the more common phylum is Firmicutes, which may vary between individuals. Another study provided evidence for individual differences in the intestinal microbiota between aging adults, depending on their health status. The composition of the intestinal microbiota of people receiving long-term residential care...
The presence of a core microbiota, dominated by symbiotic bacterial taxa (Ruminococcaceae, Lachnospiraceae and Bacteroidaceae). Moreover, in semi-supercentenarians (aged 105–109) the microbial ecosystem is enriched in Akkermansia, Bifidobacterium and Christensenellaceae – health-associated bacteria, known to promote immunomodulation and metabolic homeostasis (Biagi et al., 2016). In addition, it is also indicated that SCFAs may play an important role in the aging process. As reviewed by Riaz Rajoka and others (Riaz Rajoka et al., 2018) there is a relationship between gut microbiota and aging related diseases. Moreover, the change in composition of the gut microbiota affects production of SCFAs (mainly butyrate) which have anti-inflammatory properties, and reduction in their level may increases inflammation in elderly people (Riaz Rajoka et al., 2018).

Aside from microorganism-associated molecular patterns (MAMPs) that are expressed by components of the resident microbiota, as well as pathogens recognized by pattern recognition receptors (PRR), the microbial metabolites also influence the host’s intestinal microflora and immune system. In the process of anaerobic fermentation of exogenous components – such as dietary fiber and other indigestible carbohydrates from food – the products, i.e SCFAs, emerge that modulate the immune mechanisms and ensure immunity determined by the integrity of the mucous membrane. The intestinal mucosal barrier plays an essential role in separation of the inside of the body from the outside environment. This highly-specialized barrier composed of the mucous layer, epithelium, and lamina propria—populated by innate and adaptive immune cells – actively participates in homeostatic responses to microorganisms without causing inflammation. If, however, this barrier is broken, the mechanisms of innate and adaptive immune response induce microorganism combat and tissue repair (Maynard et al., 2012).

**SHORT-CHAIN FATTY ACIDS (SCFAs)**

Products of microbial activity, short-chain fatty acids (SCFAs) have one to six carbon atoms (C1-C6). They include acetate (C2), propionate (C3), and butyrate (C4), which are essential for intestinal homeostasis, since they maintain proper microbiome dynamics through inhibiting proliferation of some bacteria at a low pH environment (Tramontano et al., 2018). These acids affect immune reactions not only in the intestines, but also in distant tissues (Park et al., 2015; Milligan et al., 2017). Acetate emerges as a product of pyruvate conversion, either directly from acetyl-CoA or by the reductive acetyl-CoA or Wood–Ljungdahl pathway (Ragsdale & Pierce, 2008). Propionate is synthesized via the succinate pathway. Additionally, it is produced by the acylate pathway from lactate as a precursor, with simple sugars (pentoses and hexoses) as substrates in these reactions. Propionate is also synthetized via the propanediol pathway with deoxyhexoses (fucose and rhamnose) as substrates (Reichardt et al., 2014). Butyrate is formed by classical routes as a result of the reduction of acetoacetyl-CoA to butyryl-CoA, converted to butyrate by transbutyrylase and butyrate kinase (Pryde et al., 2002). Butyryl-CoA may yield butyrate via butyryl-CoA transferase-acetate Co-A (Duncan et al., 2002; Vidal et al., 2014) (Table 1). These acids are transported to intestinal epithelial cells via simple diffusion and active transport. C2 and C3 are absorbed by blood and distributed to other tissues and organs, whereas C4 is the source of energy and influences me-
Table. 1 Biosynthesis of SCFAs in the intestine

<table>
<thead>
<tr>
<th>SCFA</th>
<th>Biosynthesis</th>
<th>Microorganisms</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>acetate</td>
<td>from pyruvate in acetyl-CoA pathway</td>
<td>Akkermansia muciniphila, Bacteroides spp, Bifidobacterium spp, Prevotella spp, Ruminococcus spp.</td>
<td>Koh et al., 2016</td>
</tr>
<tr>
<td>succinate pathway</td>
<td>Blautia hydrogenotricha, Clostridium spp, Streptococcus spp.</td>
<td>Blautia hydrogenotricha, Clostridium spp, Streptococcus spp.</td>
<td>Koh et al., 2016</td>
</tr>
<tr>
<td>propionate</td>
<td>Bacteroidetes spp, Roseburia spp, Firmicutes, Roseburia insulivorans, Ruminococcus spp, Clostridium spp, Clostridiales bactrium, Eubacterium spp, Coprococcus spp, Dialister succinitiphilus, Phascolarctobacterium succinatutilens, Akkermansia muciniphila</td>
<td>Reicardt et al., 2014</td>
<td></td>
</tr>
<tr>
<td>acrylate pathway</td>
<td>Clostridium sp, Clostridiales bacterium, Coprococcus catus, Clostridium sp.</td>
<td>Reicardt et al., 2014</td>
<td></td>
</tr>
<tr>
<td>propanediol pathway</td>
<td>Roseburia insulivorans, Ruminococcus spp, Eubacterium hallii, Clostridium sp.</td>
<td>Reicardt et al., 2014</td>
<td></td>
</tr>
<tr>
<td>butyrate transferaseacetyl-Co-A pathway</td>
<td>Roseburia intestinalis, Eubacterium rectale, Roseburia insulivorans, Clostridiales bacterium, Anaerostipes hadrus, Coprococcus spp, Clostridium symbiosum, Faecalibacterium prasnitzii</td>
<td>Vital et al., 2014</td>
<td></td>
</tr>
<tr>
<td>butyrate kinase pathway</td>
<td>Bacteroidetes spp, Coprococcus spp.</td>
<td>Vital et al., 2014</td>
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RECEPTORS FOR SHORT-CHAIN FATTY ACIDS

Free fatty acids receptors (FFARs), belonging to the GPCR family, recognize SCFAs and take part in signal transmission. They are polypeptides consisting of seven transmembrane α-helix domains (7 TMD). These domains consist of an extracellular N terminus with three extracellular loops (EL1, EL2, EL3), and an intracellular C terminus with three intracellular loops (IL1, IL2, IL3). SCFAs activate four different receptors located in the cell membranes. These include mammalian FFAR2 (also known as GPR43) and FFAR3 (also known as GPR41), hydroxyarboxylic acid receptor 2 (HCA2) (also called GRP109α), as well as other receptors belonging to the GPCR family: Olfr-87 in mice, and OR51E2 in humans (Bolognini et al., 2016; Pluznick, 2016; Ohira et al., 2017). GPR43 is mainly stimulated by acetic, propionic and butyric acids. However, acids that have the greatest potential to activate GPR41 are the propionic and butyric acids, and slightly less effective – the acetic and formic acids (Brown et al., 2003; Le Poul et al., 2003). Apart from niacin and ketone bodies, the butyric and β-hydroxybutyric acids are also ligands for GPR109α. Propionic and acetic acids are produced by bacterial microflora and are also activators for Olfr-87 (Pluznick et al., 2013). What is more, SCFAs (mainly C4 and C3) are believed to take part in the activation of the peroxyisome proliferator-activated receptors γ (PPARγ), stimulating synthesis of angiopoietin-like protein 4 (ANGPTL4/FIAF) in the large intestine adenocarcinoma cells, among others (Alex et al., 2013). ANGPTL4 is a newly recognized adipokine, regulating lipid metabolism. It may also be a joining factor between the gut microflora composition and accumulation of the adipose tissue (Korek & Krauss, 2015).

The main places of GPR43 expression are digestive tract epithelial cells and the immune system cells (Brown et al., 2003; D’Souza et al., 2017). This receptor can also be found on adipocytes in the adipose tissue (Brown et al., 2003). GPR41 is present on many cells of the human body, such as the large intestinal lamina propria cells, spleen cells, lymph nodes, bone marrow, adipocytes, polymorphonuclear leukocytes, and the peripheral nervous system cells (Brown et al., 2003; Le Poul et al., 2003). Both receptors are also found on cells in the distal tubules and kidney collecting ducts (Kobayashi et al., 2017). The presence of GPR109α has been confirmed in the large intestinal epithelium, and on immune system cells: macrophages, monocytes, dendritic cells, neutrophils, and adipocytes (Brown et al., 2003), but not on lymphocytes in the white and brown adipose tissue (Koh et al., 2016). Olfr-87 mainly occurs on neurons, enterocordene cells, epithelium of the large intestine, renal arteries, and smooth muscles of blood vessels (Table 2).

SCFAs SIGNALING PATHWAY

Ligand binding and activation of the receptor initiates the signal transmission cascade in the cell. In the case of GPCR signaling pathways, the first elements 'receiving' signal from the stimulated receptor are heterotrimeric G proteins. In a resting state, G proteins exist as complexes of α, β and γ subunits. Activity of G proteins is regulated by the guanosine diphosphate (GDP)-bound α-subunit. Gα subunits include four protein subclasses: Gαs, Gαi, Gαo, Gz(Gαq), Gz γ12/13, and Gzα each of them activating a specific signaling pathway. Upon ligand binding to the receptor, GDP dissociates from Gα, which in turn binds to GTP. The Gz-GTP complex dissociates from Gα, which in turn binds to GTP. The Gz-GTP complex dissociates from Gα, which in turn binds to GTP.
**Table. 2 Receptors for short-chain fatty acids**

<table>
<thead>
<tr>
<th>Receptor</th>
<th>Location</th>
<th>SCFA</th>
<th>References</th>
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<tbody>
<tr>
<td>FFAR2 (Free fatty acid receptor 2) (GPR43)</td>
<td>digestive tract epithelial cells, immune system cells, adipocytes in adipose tissue</td>
<td>acetate, propionate, butyrate</td>
<td>Brown et al., 2003; D’Souza et al., 2017</td>
</tr>
<tr>
<td>FFAR3 (Free fatty acid receptor 3) (GPR41)</td>
<td>large intestine lamina propria cells, spleen cells, lymph nodes, bone marrow, adipocytes, polymorphonuclear leukocytes, and in peripheral nervous system cells, distal tubules and kidney collecting ducts</td>
<td>acetate, propionate, butyrate</td>
<td>Brown et al., 2003; Kobayashi et al., 2017</td>
</tr>
<tr>
<td>GPR109a (HCA2, hydroxyxycarboxylic acid receptor 2)</td>
<td>large intestinal epithelium, macrophages, monocytes, dendritic cells, neutrophils, and adipocytes</td>
<td>niacin, ketone bodies, β-hydroxybutyric acids, butyrate</td>
<td>Brown et al., 2003</td>
</tr>
<tr>
<td>Olfr-87 (murine) ORS1E2 (human)</td>
<td>neurons, enteroendocrine cells, the epithelium of the large intestine, renal arteries, smooth muscles of blood vessels</td>
<td>acetate, propionate, butyrate</td>
<td>Brown et al., 2003; D’Souza et al., 2017; Pluznick et al., 2013; Pluznick et al., 2016; Ohira et al., 2017</td>
</tr>
<tr>
<td>PPARγ</td>
<td>large intestine adenocarcinoma cells</td>
<td>propionate, butyrate</td>
<td>Alex et al., 2013</td>
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A signal from stimulated FFAR2 (GPR43) and FFAR3 (GPR41) takes place via proteins from the heterotrimeric G-protein family – $G_{i/o}$ and $G_{q/11}$ (Le Poul et al., 2003; Kobayashi et al., 2017; Milligan et al., 2017) (Fig. 1).

Signaling via FFARs plays an important part in modulation of the immune and inflammatory response. A study with mice had demonstrated that abnormalities in the expression of these receptors contribute to the development of immune cells, increase the number of Treg cells and affects the T lymphocyte and dendritic cell (DC) functions.
opment of diabetes, obesity, colitis, colon cancer, atherosclerosis, as well as hypertension and respiratory diseases (especially asthma). What is more, in vitro examination in human monocytes revealed that FFA2 and FFA3 interact to form a heteromer that may play a part in the activation of a signaling pathway with p38 protein, and this additionally contributes to immune diseases (Ang et al., 2018). Another known function of FFARs stimulated by SCFAs (mainly propionic acid) is their modulatory role in the activity of the sympathetic nervous system which helps to maintain homeostasis in the organism, which has been confirmed with a murine model (Kimura et al., 2011). As a result of GPR41 stimulation on adipocytes in the adipose tissue, C3 contributes to secretion of leptin in cultured adipocytes and in an animal model based study (Xiong et al., 2004). The murine Olfr78 receptor, on the other hand, activated by propionate, is involved in renin secretion and in regulation of blood pressure (Pluznick et al., 2013).

**SHORT-CHAIN FATTY ACIDS, POST-TRANSLATIONAL MODIFICATIONS AND THE IMMUNE RESPONSE**

SCFAs inhibit histone deacetylases (HDACs), which are involved in regulation of expression of genes promoting pathogenesis in many diseases (Grabarska et al., 2013). Overexpression of HDACs results in the reduction of histone acetylation, thus leading to gene silencing during transcription (Grabarska et al., 2013). As an example, SCFAs may act on mononuclear blood cells and neutrophils through inhibition of HDAC, which consequently reduces production of the proinflammatory tumor necrosis factor (TNF) by these cells and leads to inactivation of a transcription factor – nuclear factor kappa B (NFκB) (Rooks & Garrett, 2016).

Enzyme-controlled histone post-translational modifications (HPTMs) are a key element of gene expression regulation (Fig. 1). The best elucidated HPTM mechanism is histone acetylation, controlled by acetyltransferases (HATs) and deacetylases (HDACs) (Fellows et al., 2018). In the process of histone deacetylation, removal of the acetyl group from εN-acetyl-lysine results in a more effective DNA coiling around histones. Increased acetylation and hyperacetylation of chromatin determines its transcriptional activity, which is essential for gene expression. An impact of HDAC inhibition on the functioning of immune cells has been confirmed for murine Treg lymphocytes (Tao et al., 2007). Tao and others (Tao et al., 2007) reported that the HDAC inhibitor (HDACi) increases gene expression of Foxp3 in Tregs, and improves their suppressive properties.

It has been also found in a murine model study that the acetylation of Foxp3 promotes its binding to the Il2 promoter region, which results in the inhibition of endogenous IL-2 secretion. IL-2, on the other hand, is involved in the conversion of Foxp3+Treg cells to colitis-related Foxp3+IFN-γ+T cells (Feng et al., 2011). Another example is the impact of HDAC inhibition on maturation, differentiation, and immunogenicity of DCs (Chauviré et al., 2014; Nencioni et al., 2007). SCFAs, especially butyric acid, have been confirmed to inhibit the HDAC activity (Chang et al., 2014; Waldecker et al., 2008). The SCFA produced by commensal bacteria, butyrate – as an inhibitor of HDAC can modulate the function of macrophages present in the lamina propria of mice gut (Chang et al., 2014). A study on a mouse model provided evidence that in the mechanism dependent on HDAC inhibition, the expression of genes encoding proinflammatory mediators (nitric oxide synthase – Nos2, Il6, Il12a, and Il12b) is reduced. Lower levels of NO, IL-6, and IL-12 produced by macrophages can help to provide and maintain the immune homeostasis, and decrease inflammation in the colon (Chang et al., 2014). Currently, we can also identify other post-translational modifications, such as crotonylation, butyrylation, and hydroxybutylation. SCFAs, which are essential for normal intestinal physiology, are an important source of energy for intestinal epithelial cells and a vital factor joining the cell metabolism to translational regulation. The most recent data indicate that SCFAs are also crucial for the process of crotonylation (Fellows et al., 2018), which is a stronger activator of transcription than acetylation, and involves joining crotonyl-CoA to histone lysine residues (Sabari et al., 2015). Fellows and others (Fellows et al., 2018) analyzed crotonylation in the murine tissues of colon, brain, liver, spleen, and kidneys. This process was the most intense in the intestine, and very intense in the brain. It was also demonstrated that modification of histones depended on SCFAs, mainly butyric acid (C4), whose level was highest in the intestine, but also high in the blood serum, which explains why crotonylation is observed in distant tissues (for example, in the brain), and not only locally in the intestines. The available report (Fellows et al., 2018) suggests that SCFAs (mainly C4) increase histone crotonylation in intestinal epithelial cells through inhibition of the decrotonylase activity of HDACs. Another very important factor contributing to the process of crotonylation is the composition of the intestinal microflora that can be modified by the use of antibiotics. As shown by Fellows and others (Fellows et al., 2018), the loss of the commensal intestinal microflora substantially contributes to a decline in SCFA levels, both in the intestinal tissues and in the serum. On the other hand, SCFA deficiency leads to an increased expression of HDAC2 in the intestine and loss of histone crotonylation. Expression of HDAC2 leads to pathologies, such as tumorigenesis in the large intestine human tissue which has been confirmed by Ashktorab and others (Ashktorab et al., 2009). The latest research, but conducted with a murine model of intestinal tumorigenesis (Ravillah et al., 2014), confirmed that inhibition of HDAC2 influences inhibition of tumorigenesis through decreased expression of G2/M phase cell cycle regulatory proteins, Bel-2 protein and upregulated expression of p53 protein in the mouse colon tissue.

**IMMUNE RESPONSE MODULATED BY SHORT-CHAIN FATTY ACIDS**

SCFAs exert an impact on cells by three different mechanisms. The first of them is the activation of cellular receptors, resulting in cell proliferation or differentiation. Secondly, after penetrating into cells, SCFAs affect their metabolism. Thirdly, SCFAs act as HDAC inhibitors (Astakhova et al., 2016). In a human cell line based study, Astakhova and others (Astakhova et al., 2016) demonstrated how SCFAs influenced signaling pathways in cancerous epithelial and lymphoid cells (B-cell lymphoma, BL), which were additionally Epstein-Barr virus (EBV) positive. The SCFA that they focused on was N-acetyl butyric acid, which triggered proinflammatory IL-8 and IL-6 expression and increased NFκB activity in the cancerous epithelial and lymphoid cells. Moreover, SCFAs induced early stages of the lytic cycle of the Epstein–Barr virus (EBV), which was confirmed by expression of viral proteins involved at this stage of a viral
cycle. Butyric acid also had an effect on the induction of apoptosis in lymphoid cancerous cells. Another parameter assessed by Astakova and others (Astakova et al., 2016) was inhibition of cancerous cells with high migration potential. The authors found that even very low levels of N-acetyl butyric acid inhibited migration of these cells. They demonstrated that transport proteins MCT1 (monocarboxylate transporter 1, also known as solute carrier family 16 member 1) and MCT4 (monocarboxylate transporter 4, also known as solute carrier family 16 member 3) were essential for the SCFA activity and penetration into cells. These data show that the presence of SCFAs and their influence on transporter cells enable elimination of cancerous and EBV-infected cells (Astakova et al., 2016).

SCFAs promote differentiation of T lymphocytes into effector T cells (Te) and regulatory T cells (Treg) via HDAC inhibition but it is independent of GPR41 or GPR43. The mTOR-S6K kinase pathway is essential for differentiation of the T lymphocytes (Park et al., 2015). Inhibition of HDACs in T lymphocytes by SCFAs (C2 and C3) can enhance phosphorylation of the ribosomal protein S6 (rS6), which is the major target of the mTOR pathway. It has an impact on expression of IL-10, IL-17 and IFNγ – the key effector and regulatory cytokines in T cells. Based on in vitro research, it has been demonstrated that C2 can regulate T cell populations, which is modulated by immunological context and cytokine milieu. The population of T lymphocytes secreting IL-10 were increased by SCFA in the steady condition in vitro, whereas in an active immune response, only C2 increased the number of the effector T cells (Park et al., 2015). This phenomenon seems to be important because secretion of IL-10 by Te lymphocytes determines their anti-inflammatory properties, which is critical in weakening the immune response in inflammatory diseases of various organs. These research also suggested that propionate in the intestine may promote the differentiation of Treg cells producing IL-10 and expressing the Foxp3 gene. What is more, both C2 and C3 (in vitro) promote differentiation of naive T lymphocytes into T helper 17 (Th17) cells, and in the presence of IL-12 they lead to induction of Th1 lymphocytes. This, however, depends on the dose and the level of the acid. Th1 and Th17 lymphocytes are involved in the immune response to pathogens, and mediate the inflammatory response (Park et al., 2015).

SCFAs AND THE URINARY TRACT

A favorable effect of SCFAs is well known. There are data, however, indicating that their permanently elevated levels contribute to inflammatory states, which was demonstrated with the example of the murine urinary tract model (Park et al., 2016). Long-term administration of SCFAs (especially C2) was found to cause their substantial increase in the intestinal tissue and blood, with the highest level observed in the renal tissues. Immunohistochemical analysis confirmed that mice with chronic renal disease (CRD) provided with C2, had elevated tissue levels of CD4+ T and CD11c+ lymphocytes. An increase was also observed in the levels of Th1 and Th17 effector lymphocytes, as well as IL10+ and Foxp3+ T cells. Additionally, in the tissues of the animals with CRD, the researchers observed a considerably higher expression of mRNA for IL-6, IL-17, IFNγ, CCL2, CCL5, CCL17, as well as for IL-10 and TGFβ-1. The results reported by Park and others (Park et al., 2016) suggest that irrespective of the presence of GPR41 and GPR43, as a result of activation of the mTOR signaling pathway, SCFAs promote differentiation of T lymphocytes into Th1 and Th17, which seems to be critical for the occurrence of not severe but chronic inflammatory states in the CRD pathogenesis. A long-lasting increase in SCFA levels in the body is a consequence of dysbiosis. Through absorption of SCFAs into the blood circulation system and activation of signaling pathways in the immune system cells, such a state induces chronic inflammation which can lead to tissue damage in various organs (kidneys, for example) (Park et al., 2016). In spite of this, favorable effects of SCFAs (mainly the acetic and butyric acids) have been demonstrated in human population based studies when restraining the oxidative stress and inflammatory response in renal glomerular mesangial cells (GMC). Aside from a decrease in proliferation of GMC, as well as substantial inhibition of reactive oxygen species (ROS) and malondialdehyde (MDA), increased levels of an antioxidant enzyme – superoxide dismutase (SOD) – were observed. What is more, GMC stimulated by high levels of glucose and bacterial lipopolysaccharides (LPS) in the presence of SCFAs, secrete a substantially lower amount of IL-1β proinflammatory cytokine and leukocyte chemotactic factor, called CCL2 chemokine or monocyte chemotactic protein (MCP-1) (Huang et al., 2017). This suggests that maintaining normal levels of SCFAs and identification of signaling pathways in which SCFAs are involved, can be essential for reducing inflammation in the renal diseases. Research on SCFAs as factors potentially reducing inflammatory states and fibrosis in renal tissues was also conducted by Kobayashi and others (Kobayashi et al., 2017). They noticed that propionic acid in the human renal cortical epithelial cells (HRCE) decreases MCP-1 expression dependent on TNF-α. This mechanism is determined by lower phosphorylation of the p38 protein and the c-Jun N-terminal kinase (JNK) via the pathway dependent on GPR41 and GPR43, which in this case joins mainly with the Gβγ proteins (Kobayashi et al., 2017).

SCFAs AND THE CENTRAL NERVOUS AND RESPIRATORY SYSTEMS

Dysregulation of intestinal microbiota composition, and thus its metabolites, can also affect autoimmune reactions, including those in the central nervous system (CNS). Moser and others (Moser et al., 2017) analyzed patients with the clinically isolated syndrome (CIS) and patients with the early phase of multiple sclerosis (MS). The term CIS refers to a first episode of neurologic symptom onset of potential multiple sclerosis (MS). It is caused by inflammation and loss of myelin covering the neurons (demyelination) in the CNS (Efendi, 2016). Moser and others (Moser et al., 2017) noticed that the levels of acetic and butyric acids in feces samples of the CIS and MS patients were significantly lower than in the control group. Furthermore, the patients with demyelinating disease had considerably fewer CD103+ tolerogenic dendritic cells (DC) and CD4+25+127- Treg lymphocytes in the descending colon bioplates than the control group. These data suggest a relationship between a decline in the SCFA levels and deficiency of DCs and Treg cells stimulated by these acids, and pathological symptoms in MS patients (Moser et al., 2017).

The influence of intestinal dysbiosis and disturbed SCFA levels have been also confirmed as factors exacerbating inflammation in allergies of the respiratory
system (lungs) (Cait et al., 2017). Research with a murine model has provided evidence that antibiotic (vancomycin) treatment reduces both, the number and variety of intestinal bacteria species, producing SCFAs. It is worth mentioning that mice receiving vancomycin exhibited an enhanced allergic reaction, confirmed by an increase in eosinophils in the airways, and elevated serum IgE levels. An increased immune response was inhibited in animals by butyric acid (C4) supplementation. In the same study, in vitro examination of cells taken from mice receiving antibiotics revealed that C4 decreases population of the CD4+ T lymphocytes in which expression of IL-4 is observed. IL-4 is mainly produced by stimulated Th2 lymphocytes, and takes part in the change of antibody classes from IgM through IgG to IgE. This suggests that there are relationships between the activation of Th2 cells, increased expression of IL-4, elevated serum IgE levels, and an inflammatory allergic reaction. Also, Cait and others (Cait et al., 2017) analyzed the anti-inflammatory influence of SCFAs on murine DC cells. Genetic analysis had shown that DC cells activated by contact with an allergen, but subjected to C4 activity, were less likely to stimulate T lymphocytes and their chemotactic properties were also weaker. These data imply that intestinal dysbiosis increases an allergic reaction in lungs by stimulating inflammatory response mechanisms in which T lymphocytes and DC cells are involved. Nestasi and others (Nastasi et al., 2015), who performed in vitro research to assess modulatory influence of SCFAs on the function of human monocyte-derived dendritic cells isolated from peripheral blood, demonstrated that GPR41 and GPR109A expression was like in the primary human CD1c+ and CD141+ DC cells. In the same study, they also found that SCFAs that play the most important part in modulation of the function of mature dendritic cells are the propionic and butyric acids, reducing secretion of proinflammatory chemokines (among them, CCL3, -4, -5 and CXCL9, -10, -11, but not CCL2, -11, and CXCL5). Both, C3 and C4 substantially reduced expression of IL-6 and IL-12B—proinflammatory cytokines induced by bacterial LPS. The results of that study had clearly shown that propionic and butyric acids are those SCFAs that have the greatest immunomodulatory and direct influence on mature dendritic cells, which was manifested as inhibition of gene expression for selected cytokines and chemokines (Nastasi et al., 2015).

SCFAs AND A DISTAL EFFECT – AN IMMUNE REACTION IN THE EYE

Distant effects of SCFAs were demonstrated with the example of experimental autoimmune uveitis (EAU) in a murine model by Nakamura and others (Nakamura et al., 2017). The authors had confirmed a decreased length of the muscular layer of the small intestine and intestinal crypts in immunized mice, but this morphological change was significant during the beginning of uveitis. In propionate treated EAU mice the depth of intestinal crypts and villi length was increased. Moreover, oral administration of SCFAs changed the frequency of Treg cells in the cecal and colonic lamina propria lymphocytes (LPL), cervical lymph nodes (CLN), mesenteric lymph nodes and spleen, depending on time after administration SCFAs. Among others, SCFA supplementation given to the animals had alleviated the eye inflammation by induction of Treg cells, both in the intestines and the distant tissues and lymphatic organs. The researchers had noticed that administration of SCFAs abates migration of Th1 lymphocytes from the intestines to the spleen (Nakamura et al., 2017). Additionally, the connection between the intestinal homeostasis and uveitis may explains why the latter accompanies the autoimmune inflammatory diseases (Nakamura et al., 2017).

SCFAs AND THE IMMUNE REACTION IN OBESITY

The metabolism of SCFAs is also a factor contributing to obesity. There are considerable differences in the microflora composition between slim, overweight, and obese people. Overweight and obese individuals have more Bacteroidetes mediating the production of propionic acid, which after absorption and distribution to the liver can become a substrate in the processes of glucogenicosis or lipogenicosis. Obese people have noticeably higher C3 levels than slim ones (from 84.6 mmol/l to 103.87 mmol/l) (Schwierz et al., 2010). There are also data indicating that both, too high and too low SCFAs levels can lead to obesity and other symptoms of the metabolic syndrome. A murine model research provided evidence that a long-term increase in C2 levels results in insulin resistance and elevated levels of triglycerides in the plasma, liver, and muscles (Perry et al., 2016). Excessive adipose tissue, progressive obesity, and the loss of metabolic homeostasis are the causes of inflammatory states. The main metabolic organ regulating energy balance is white adipose tissue (WAT) composed of adipocytes and stromal tissue which contains immunological cells. Immune system of the adipose tissue consists of M2 macrophages (Mf), type 2 innate lymphoid cells (II.C2), invariant natural killer T (iNKT) cells, as well as Th2 and Treg lymphocytes. Macrophages are cells that contribute to an inflammatory response associated with adipose tissue and stronger insulin resistance. Based on their experimental murine model, Nakajima and others (Nakajima et al., 2017) analyzed the impact exerted by SCFAs on the activation of GPR43 in the adipose tissue macrophages. What is interesting, the activated receptor fulfills different functions in the M1-type and M2-type macrophages. In the GPR43-dependent mechanism, TNFα expression is observed in M2 macrophages, but not in M1 macrophages. It shows that in the state of equilibrium in the adipose tissue, GPR43 regulates the SCFA-dependent immune response. TNFα, secreted by anti-inflammatory Mf with M2 phenotype, may contribute to suppression of adipose tissue accumulation by lowering the expression of glucose transporter isof orm 4 (GLUT4) in the adipose tissue (Nakajima et al., 2017). Similar evidence for the role of SCFA-stimulated GPR43 and GPR41 in prevention of gaining weight and inhibition of obesity-related chronic inflammatory states was reported by Lu and others (Lu et al., 2016) in a murine model study. The authors also noticed a relationship between stimulated receptors and the activation of the processes of cellular metabolism — adipogenesis and oxidation. It should be emphasized that obese animals had elevated levels of proinflammatory IL-6, IL-1β, and MCP-1, which were substantially reduced by means of SCFA supplementation (Lu et al., 2016). Additionally, abnormal intestinal microflora was proved to increase the levels of SCFAs in feces and IL-6 in serum, which has an impact on lipid metabolism and the occurrence of subclinical metabolic changes in people (Rodriguez-Carrio et al., 2017).
**DYSBIOSIS, SHORT-CHAIN FATTY ACIDS AND INFLAMMASOMES**

Effects of SCFAs are locally observed in the intestines. Recent data shows that SCFAs are bacterial metabolites indirectly involved in the activation of inflammasomes (Fig. 1). Inflammasomes are known as multi-protein cellular complex composed of a NOD-like receptor protein, the adaptor protein ASC and pro-caspase-1. Induction of this cellular complex leads to initiation of assignaling cascade in which activated caspase-1 contributes to secretion of interleukin IL-1β and IL-18 (Broz & Dixit, 2016). IL-1β belongs to the IL-1 interleukin sub-family, its role in intestinal immunity has been reviewed by (Manzaillon et al., 2015). In intestinal immunity, IL-1β may have both, a protective and pathogenic function. Murine model studies demonstrated that genetic deletion of IL-1β retards the recovery of a gut epithelium. Moreover, IL-1β can modulate expression of other cytokines i.e. IL-22 in Th17 and Th22 cells. What is more, IL-1β may affect Treg cell differentiation and contribute to inflammatory state by inducing the Th17 cell differentiation. In synergy with TNF-α, IL-1β may play a role as an enhancer of epithelial tissue damage (Manzaillon et al., 2015). In a human pneumocystis cell line (THP-1 cells) (Ohira et al., 2012) investigated the effect of SCFA – butyrate, on expression of IL-1β. Their study suggests that butyrate (in LPS stimulated THP-1 cells) enhances IL-1β production by activating caspase-1 via reactive oxygen species, as well as phosphorylation of the MAP kinase pathways and G protein mediated pathways (Ohira et al., 2012). These data indicate that butyrate may play an important role in elevation of the inflammatory state. IL-18 is believed to be a cytokine that takes part in maintaining homeostasis and integrity, as well as the reconstruction of the intestinal epithelium. As shown with a murine model, a high-fiber diet, and thus increased SCFA production by intestinal bacteria, causes enhanced activation of the NLRP3 inflammasome in intestinal epithelial cells, as manifested by an increase in the IL-18 levels. Signaling mechanisms of inflammasome activation involve GPR43 and GPR109A, which cause hyperpolarization and an increased inflow of Ca2+ ions (Macia et al., 2015). Aside from fiber metabolites, a factor that activates inflammasome in the intestines is the bacterial flora, whose composition and diversity depends on the type and intake of dietary fiber. In the study of Macia et al. (Macia et al., 2015), the microflora isolated from mice consuming a greater amount of dietary fiber substantially increased the release of the active form of caspase-1 which clearly indicates enhanced induction of the inflammasome. Also, Levy and others (Levy et al., 2015) carried out research on the role of the intestinal microflora in inflammasome-related mechanisms, based on the murine model. They found that certain substances, such as taurine, histamine, and spermine, indirectly regulate production of intestinal antimicrobial peptides (AMPs) (interleukin 1 (ITLN1), resistin-like molecule β (RELMβ), angiogenin) through stimulation of NLRP6 inflammasomes and secretion of IL-18. Distortion of AMP balance by inflammasome deficiency drives dysbiosis development. Commensal bacteria take part in transcription of inflammasome components, and probably induce inflammasome formation, thus affecting IL-18 secretion. The authors also informed that the NLRP6/IL-18/AMP signaling pathway ensures normal bacterial composition both, at the level of alpha- and beta-diversity (Levy et al., 2015). Nevertheless, Mamantopoulos and others (Mamantopoulos et al., 2017), had denied the NLRP6 inflammasome to participate in the modulation of the gut microflora composition in a mice based study. Also, the AIM2 inflammasome, regulating the IL-18/IL-22BP/IL-22 signaling pathway, promotes the maintenance of intestinal homeostasis and decrease in dysbiosis which has been confirmed with a murine model. In response to bacterial DNA, an active AIM2 inflammasome mediates IL-18 secretion, reducing IL-22BP (IL-22 binding protein) expression, and thus leading to an increased activity of IL-22 in the intestine. Under such conditions, normal secretion of AMPs (such as REG3β and REG3γ) occurs to control the composition of the intestinal microbiota, and to prevent dysbiosis. In the contrast, loss of AIM2 signaling predisposes to intestinal inflammation associated with abnormal functioning of the intestinal microflora. This is because reduced secretion of IL-18 boosts expression of IL-22BP, and decreases secretion of IL-22 and AMPs. During intestinal inflammation, on the other hand, elevated IL-18 levels reduce IL22BP expression via the IL-22 pathway, and thus activate STAT3 signaling pathways which increase the levels of REG3β and REG3γ. Increased AMP production re-stimulates STAT3 based on a positive feedback loop, which in turn affects cell proliferation (Ratsimandresy et al., 2017). Furthermore, inflammasomes and SCFAs contribute to changes in the arterial wall structure in atherosclerosis. A murine model study (Yuan et al., 2018) demonstrated different effects of various SCFAs on activation of the NLRP3 inflammasome. It had shown that butyric acid can inhibit O2− and the NLRP3 inflammasome formation and thus reduce the inflammatory response, preventing one of the causes of atherosclerosis—neointima (Yuan et al., 2018).

**CONCLUDING REMARKS**

Currently, many studies are being conducted to assess the contribution of intestinal microflora metabolites to the organism’s homeostasis. The vast majority of them concern the influence that SCFAs exert not only on the intestinal environment, but also on distant tissues and organs. Increasingly, SCFAs are believed to play an immunomodulatory role, but nonetheless the mechanisms of their activity require further investigation. They seem to be essential for the differentiation of T lymphocytes, the reduced secretion of some inflammatory cytokines, and indirect inflammasome activation. Signal sending mechanisms from stimulated FFAR receptors, for which SCFAs are ligands, are also important. At present, both, the signaling pathways and receptors, are the new therapeutic targets in many inflammatory dysbiosis-related diseases. More in-depth SCFA analysis with regard to particular diseases would certainly help us understand their etiology and pathogenesis.

**REFERENCES**


Vol. 66  Immunomodulatory potential of gut microbiome-derived short-chain fatty acids 11


Ratsimandresy RA, Indrumohan M, Dörfler A, Stehlik C (2017) The AIME 201715: pathogenicity of enterotoxigenic Bacteroides fragilis is related to intestinal homeostasis through the IL-1β/IL-22/STAT3 pathway. *Cell Mol Immunol*** **14**:127–142. [https://doi.org/10.1038/cmi.2016.35](https://doi.org/10.1038/cmi.2016.35)


Tao R, de Zoetien EF, Özkanay E, Chen C, Wang L, Porrett PM, Li B, Turkla LA, Olson EN, Greene MI, Wells AD, Hancock WW (2007) Decaycetylase inhibition promotes the generation and function of regulatory T cells. *Nat Med*** **13**:1299–1307. [https://doi.org/10.1038/nm1652](https://doi.org/10.1038/nm1652)


